News & Views

Reactive Oxygen Species Drives Myocardial Angiogenesis?

NILANJANA MAULIK

ABSTRACT

Neovascularization, the natural physiological process of formation of new blood vessels, is extremely important for ameliorating the function of the heart that undergoes ischemic stress. This process is potentially important for the treatment of ischemic heart and limb diseases, which includes formation of capillaries (angiogenesis) and collateral arteries. Ischemia or coronary artery occlusion induces vascular endothelial growth factor (VEGF) in the experimental rat myocardial infarction model, and this molecule encourages development of coronary collateral circulation and retention of the blood supply to the ischemic area. Restoration of the blood supply to the ischemic area prevents cardiomyocyte death and cardiac remodeling. Among the various triggers and enhancers of angiogenesis, hypoxic or ischemic preconditioning, as well as pharmacologic agents such as statin and resveratrol, have been identified as important stimuli for the induction of new vessel growth. It has already been demonstrated that the VEGF family and its receptor system is the fundamental regulator in the redox cell signaling of angiogenesis. This review article will focus on the role of reactive oxygen species in the process of myocardial angiogenesis. Antioxid. Redox Signal. 8, 2161–2168.

INTRODUCTION

HE CENTRAL ROLE OF VASCULAR ENDOTHELIAL growth factor (VEGF) in angiogenesis in health and disease makes it attractive as a therapeutic target for anti-angiogenic drugs and as a pro-angiogenic cytokine for the treatment of ischemic heart disease. Whereas VEGF binds to two receptor protein tyrosine kinases, VEGFR1 (Flt-1) and VEGFR2 (Flk-1/KDR), most biological functions of VEGF are mediated by Flk-1. However, studies from our laboratory have already confirmed the significance of Flt-1 receptors also in ischemic preconditioned-mediated cardioprotection (17) involving downregulation of several important genes in Flt-1knockout mice (Flt-1-/-) in an ischemic reperfused model (Table 1). The exact mechanism of VEGF signaling via Flt-1 is yet to be determined and requires well-designed studies. Recently, we demonstrated for the first time the ability of ischemic preconditioning (IPC) to induce angiogenesis in infarcted myocardium along with the activation of several transcription factors such as stat3, Pax-5, NF-kB, TFIID, SP1, and reduction of VEGF-mediated vascular permeability by

inhibition of c-src in ischemic preconditioned myocardium, thereby reducing ischemic injury in the rat myocardial infarction (MI) model (17). However, the mechanism by which activation of VEGFRs elicits these cellular events is not fully understood.

Reactive oxygen species (ROS) stimulate angiogenic responses in ischemic reperfused hearts. Several sources of ROS have been identified in the cell, the most significant one being the mitochondrial respiratory chain. In the cytosol, other intracellular sources of ROS include xanthine oxidase and cytochrome P450, as well as membrane NADPH oxidase and nitric oxide synthase (40, 60). We have already documented that in vitro short exposure to hypoxia/reoxygenation, either directly or indirectly, produces ROS in human coronary arteriolar endothelial cells that induce oxidative stress, which is associated with increased VEGF and angiogenesis or tuborogenesis (71). It is well documented that ROS can both cause tissue injury and promote tissue repair by promoting angiogenesis. Reactive oxygen species derived from gp91phox (NOX2)containing NAD(P)H oxidase are involved in angiogenesis in mouse sponge models, as well as in VEGF signaling in

Molecular Cardiology and Angiogenesis Laboratory, Department of Surgery, University of Connecticut Medical Center, Farmington, Connecticut.

Table 1. Microarray Analysis Demonstrating Significant Number of Downregulated Myocardial Genes in Flt-1-/+ Mice

Gene name	Ratio (Flt-1+/-/WT)
GRO1 oncogene (Gro1)	0.13
Exportin 4	0.16
V kappa 21–11	0.23
Anxa7	0.24
ΙΚΚβ	0.27
Human zinc finger protein (ZNF139)	0.28
CSF-1	0.29
Ubiquitin-conjugating enzyme	0.32
MgcRacGAP mRNA for GTPase activating protein	0.38
Platelet derived growth factor, alpha (PDG)	Fα) 0.41
Trinucleotide repeat containing 7 (TNRC7)	0.42
Asparagine synthetase mRNA	0.43
H3051H08	0.44
H3112H12	0.44
Zinc finger protein 57 (ZFP57)	0.47
Interferon gamma induced GTPase	0.49
Heat shock transcription factor 2 binding protein (HSF2BP)	0.52
Inducible 6-phosphofructo-2-kinase mRNA	0.52
HSP84 kDa	0.57
HSP70 kDa	0.60
JAK2	0.63
Beta-2 microglobulin (B2m)	0.63

cultured endothelial cells (16). The role of gp91phox-derived ROS in neovascularization in response to tissue ischemia is not well studied, although impairment of neovascularization recently has been demonstrated in gp91phox -/- mice with hindlimb ischemia (62). There are also no reports describing which pathways are predominant, as study of the role of ROS in angiogenesis is relatively new. We think the important pathway will depend on the relative contribution of Flk-1-dependent pathways activated after ischemic/hypoxic stress followed by reperfusion. In addition, there may be additional pathways that are Flk-1 dependent but ROS independent.

ROLE OF REACTIVE OXYGEN SPECIES IN ANGIOGENESIS

Oxygen homeostasis is of critical importance for maintaining the viability of all tissues. Lack of sufficient tissue oxygenation is predominantly caused by impaired blood flow. Attempts to restore normal oxygen levels after episodes of hypoxia or ischemia result in the generation of various types of free radicals, such as superoxide anions, hydrogen peroxide, and hydroxyl radical, collectively known as reactive oxygen species (ROS). Although lack of adequate oxygen (e.g., hypoxia) is an initiator of various diseases, it also can trigger a unique "repair" mechanism, that acts as an important inducer of angiogenesis. ROS play a very important role in signaling pathways stimulated by growth factors in vascular

cells. Recent reports suggest that ROS, such as superoxide anions (O_2 -), play an important role in mediating signals initiated by growth factors and inflammatory cytokines (11). In this regard, we have previously shown that hypoxic preconditioning (hypoxia/reoxygenation) mediates the activation of NF- κ B in rat myocardium and human coronary arteriolar endothelial cells (38, 39, 50, 54, 71)

Reactive oxygen species such as superoxide anion (O₂-•) and hydrogen peroxide (H₂O₂) are involved in the signaling pathways mediating many stress and growth responses, including angiogenesis (31). In endothelial cells, H₂O₂ stimulates cell migration and proliferation (43). Hypoxia/reoxygenation, which produces ROS, elicits capillary tube formation in human coronary arteriolar cells (14). In vivo, elevated oxidative stress directly correlates with neovascularization and VEGF expression in the retina of diabetics (49) and in aortic plaque of models of atherosclerosis (64). Hypoxia is a strong inducer for VEGF expression both in vivo and in vitro (20). Fibroblast growth factor (FGF) mRNA is increased also in the brains of animals exposed to hypoxia (10). H₂O₂-mediated strong VEGF gene expression was also demonstrated in the rat-derived endothelial cells (19). Several metabolic pathways in mammalian cells, such as xanthine/xanthine oxidase, NADPH oxidase, mitochondrial and microsomal electron transport chains, and cyclooxygenase pathway can generate ROS and H₂O₂. ROS and H₂O₂ at high concentrations are highly toxic to cells. However, at low concentrations, ROS play various physiological roles such as regulating gene expression and cell proliferation. They are also pivotal as an intracellular second messenger. H2O2 was found to induce VEGF mRNA in a dose-dependent protein kinase c (PKC)-dependent manner (56). Moreover, Shih et al. (56) suggested that PKC upregulates VEGF mRNA in human glioblastoma cells through the stabilization of VEGF mRNA. The temporal change of VEGF mRNA (28) supports the hypothesis because ischemic preconditioning (IPC) rapidly and greatly upregulated VEGF mRNA at 3-12 h after infarction, whereas there was a smaller extent of VEGF mRNA induction irrespective of the IPC procedure at 1-3 days after infarction. This report also showed that IPC might enhance VEGF gene expression and angiogenesis through nuclear translocation of PKC ϵ in the infarcted myocardium.

VEGF was rapidly induced by transient ischemia in the heart. A single episode of ischemia by abrupt ligation of a coronary artery led to a prolonged activation of VEGF mRNA in the rat heart. After 1 h of ligation, expression of VEGF mRNA, as determined by quantitative Northern analysis of the entire heart, increased by 2.5-fold. Expression of Flk-1 (VEGFR-2) followed a similar pattern, reaching a peak after a 3.7-fold increase at 1 h and then reduced slowly. Flt-1 (VEGFR-1) also increased following left anterior descending (LAD) coronary artery ligation (25). Since increased production of ROS has been well documented in myocardial ischemia and reperfusion, it is possible that such upregulation of VEGF is mediated by ROS. Several studies demonstrated the presence of AP-1 and NF-κB binding sites in the promoters of human and mouse VEGF genes. In stimulated endothelial cells, the oncogenic transcription factors c-ets-1 and c-rel/NF-kB function to induce specific gene expression (58). In these activated endothelial cells, c-ets-1

regulates the expression of a number of genes involved in matrix degradation, including collagenase, stromelysin, and urokinase-type plasminogen activator.

Reactive oxygen species derived from NAD(P)H oxidase are critically important in many aspects of vascular cell regulation, and both the small GTPase Rac 1 and gp91phox are critical components of the endothelial NAD(P)H oxidase complex. A major source of endothelial O₂-• generation is NAD(P)H oxidase, which consists of a plasma membrane spanning flavocytochrome b558 composed of gp91phox and p22phox, and cytosolic components p47phox and p67phox. The small molecular weight G protein Rac is also necessary for assembly of the active NAD(P)H oxidase complex (46). Recently it was demonstrated that a gp91^{phox} containing NAD(P)H oxidase was a major source of ROS in vascular endothelial cells (4), and that Rac 1 are critical components of endothelial NAD(P)H oxidase. In vitro studies suggest that ROS derived from gp91phox containing NAD(P)H oxidase are important in VEGF signaling and angiogenesis. More elaborate study is necessary to document the components of NAD(P)H oxidase as potential targets for angiogenic therapy in ischemic heart disease. VEGF-induced cell signaling and angiogenesis are tightly controlled by the reduction/oxidation environment at the level of VEGF receptors and will provide novel insights into NAD(P)H oxidase as a potential therapeutic target for myocardial angiogenesis. Our results in the future may suggest that ROS derived from gp91phox containing NAD(P)H oxidase are important in VEGF signaling after ischemia-reperfusion to initiate the repair mechanism followed by angiogenesis.

VEGF SYSTEM DURING ISCHEMIC STRESS

The process of angiogenesis is regulated by signals obtained from the transmembrane receptor tyrosine kinases (RTKs) and nonreceptor tyrosine kinases of endothelial cells. Flk-1/KDR and Flt-1 are two such RTKs, which, together with their ligand VEGF, have been shown to control blood vessel development during embryogenesis (55, 59). This receptor/ ligand system augments neovascularization (3, 5, 23, 32, 43, 51). Studies in rat myocardial infarction models also demonstrated significant induction of VEGF (275%), Flk-1 (375%), and Flt-1 (400%) mRNA expression throughout the entire heart after 1 h of infarction (34). A great deal of attention has been directed toward studies of VEGF expression and its function in myocardial ischemia/hypoxia (6, 21, 22, 27, 47, 48) and relatively little is known regarding the mechanism of its receptors, Flk-1 and Flt-1. VEGF is the only known ligand for Flk-1, whereas Flt-1 is able to bind to placental growth factor in addition to VEGF.

Embryological studies have demonstrated abundance of Flk-1 in human lung tissues, while Flt-1 was found to be abundant in the heart, lung, and kidneys (27). Careful investigations demonstrated functional differences between Flk-1 and Flt-1 in endothelial cells. In the developing human heart, both receptors were expressed in myocardial capillaries, and stimulated intracellular calcium flux along with VEGF stimu-

lation. Genetically manipulated Flk-1 knockout (homozygous) studies demonstrated early embryonic death due to inhibition of vasculogenesis, whereas in another study, homozygous Flt-1 disruption caused failure to assemble normal formation of vascular channels (55). In another important observation it was found that Flt-1 was expressed in endothelium of both large and small vessels, whereas Flk-1 expression was restricted to only small vessels (34). One of our recent studies demonstrated a myocardial distribution pattern of Flk-1 and Flt-1 after rats were exposed to whole body hypoxia followed by 24 h of reoxygenation. Intense staining was observed along the capillaries, in addition to strong localization around the coronary arteries (55). We have also documented the intensity of staining for both the receptors, which increased significantly in the hypoxia/reoxygenation group compared to the corresponding normoxic control group (55). Western blot analysis also documented similar results. In other words, in a systemic whole body hypoxic rat model, we found significant abundance of Flk-1 protein expression in the 1 h hypoxia group, that remained elevated in the 2 h group (hypoxic), but was reduced in the 4 h hypoxic group, followed by 24 h reoxygenation, suggesting a role in the initial rather than the later stages of the early angiogenic process. In contrast, induction of Flt-1 protein expression was increased in the 1 h hypoxia group and continued to be elevated even in the 4 h hypoxia group, indicating a more continuous role in the early angiogenic process (55). We have also documented significant improvements in myocardial function along with increased capillary and arteriolar density following induction of survival factors VEGF, Bcl-2, and survivin in the setting of the fully established chronic rat myocardial infarction model subjected to ischemic preconditioning (17, 37). Mammalian hearts can be adapted to ischemia by repeatedly subjecting it to short-term reversible ischemia, each followed by short duration of reperfusion. This phenomenon is known as "ischemic preconditioning (IPC)" (33). Substantial evidence exists to support the notion that oxygen-derived free radicals are generated during reperfusion of ischemic myocardium, resulting in development of oxidative stress.

Ischemia induces the angiopoietin/Tie receptor system in a focal cerebral ischemia model (7). Myocardial adaptation to ischemic stress in stunned pig myocardium demonstrated the induction of c-jun, c-fos, Egr-1, and jun-B that may be involved in the repair process of angiogenesis (15). Adenosine is known to limit the degree of vascular injury during ischemia and reperfusion by inhibition of oxygen free radical release that prevents endothelial cell damage. This may help preserve endothelial cell function and microvascular perfusion (29). Recently we demonstrated that in vivo brief repetitive cycles of coronary artery occlusion (5 min), followed by short duration of reperfusion (10 min), triggered myocardial angiogenesis at the capillary and arteriolar levels, which corroborated the improved myocardial contractile function (17). We concluded that ischemic preconditioning-mediated VEGF expression is cardioprotective and VEGF triggers its cardioprotective signal via its receptors Flt-1 and Flk-1 (Fig. 1). However, to date, there has been relatively little information regarding physiological control of angiogenesis by these two VEGF receptors during myocardial protection. Two other angiogenic factors, the angiopoietins 1 and 2 (Ang-1 and Ang-2),

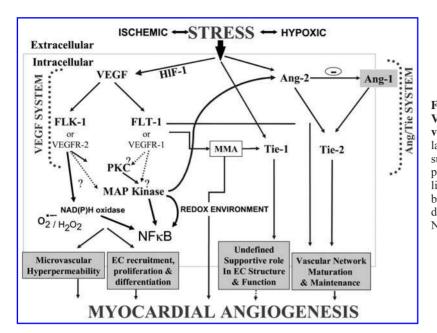


FIG. 1. Schematic diagram showing VEGF and angiopoietin-Tie system involved in cardioprotection. The stimulation of myocardial angiogenesis by stress such as ischemic/hypoxia may constitute a potential basis for a possible more long-lived adaptive response to stress afforded by preconditioning stimuli by upregulating downstream signaling molecule (such as NADPH, PKC, MAP kinases, and NF-κB).

regulate the maturation of new blood vessels from proliferated endothelial cells (69). Tie-1 and Tie-2 comprise another family of endothelial specific receptor tyrosine kinases, Angland Ang-2 being the specific ligands for Tie-2.

VEGF-mediated angiogenesis signaling is widely accepted; however, relatively little is known regarding VEGFmediated downstream signaling through Flt-1 and/or Flk-1. Evidence suggests that these two tyrosine kinase receptors of VEGF likely serve different functions in endothelial cells. These two receptors were responsible for stimulating intracellular calcium flux and were also found to be phosphorylated in response to VEGF (61). It is likely that both receptors play a significant role in VEGF-mediated angiogenesis in several pathological as well as physiological situations. Thus, we were very interested in documenting the candidate genes involved in VEGF-mediated signaling through its receptors Flk-1 and Flt-1 in ischemic preconditioned-mediated angiogenesis in an MI model. As we know from literature and also from our own work, VEGF, Flk-1, and Flt-1 expression is linked with angiogenesis and expression of the growth factors, and the same receptors cannot initiate angiogenesis without additional factors. In our study, PC-regulated VEGF-mediated cardioprotection in the WT mice determined several signaling molecules that are involved in VEGF-mediated signaling through Flt-1 receptors. There was no inhibition of VEGF expression, as shown in the Flt-1+/- knockout mice, and the extent of the expression of VEGF was found to be the same in both the wild type and Flt-1+/- knockout with the PC group (1). We have demonstrated clearly that NF-κB plays a significant role in angiogenesis and that inhibition of this redox molecule inhibits the capillary and arteriolar density and tuborogenesis in vivo and in vitro (51, 71). Real time PCR for CSF-1 also demonstrated significant involvement of PC-mediated cardioprotection in wild-type control compared to Flt-1^{+/-} knockout. CSF-1 binding to its receptor CSF-1R on cells results in tyrosine phosphorylation of the receptor and many

other proteins. It plays a significant role in cell survival and cell proliferation. In addition, significant downregulation of CSF was reported in the development of heart failure after MI (70). Therefore, our result demonstrates that VEGF-mediated signaling through Flt-1+/- involves several factors such as NF-κB, CSF-1, HSP 70, and HSP 84. Preconditioning mediated cardioprotection was found to be significantly affected in Flt-1+/- knockout mice as observed by the functional recovery. The functional recovery was not the same in Flt-1+/as found in the wild type when subjected to PC (1). The infarct size was found to be significantly higher in the preconditioned Flt-1+/- PC myocardium when compared to the wildtype PC control. The microarray as well as real time PCR data demonstrated significant downregulation of JAK2 mRNA in Flt-1^{+/-} knockout myocardium after ischemia and reperfusion (1). Therefore, in our study we found disruption of cardioprotection in Flt-1^{+/-} KO mice when compared to the wild type.

NITRIC OXIDE AND VEGF EXPRESSION

NO significantly contributes to the prosurvival/proangiogenic process of capillary formation and maturation by triggering and transducing cell growth and differentiation via endothelial-constitutive NO synthase (ec-NOS) activation, cyclic GMP (cGMP) elevation, mitogen activated kinase (MAPK) activation, and fibroblast growth factor-2 (FGF-2) expression (13). Nitric oxide constantly accompanies O₂ and/or ROS in most of their physiological activities. NO is a unique messenger in that it is produced in one cell and diffuses into adjacent target cells to activate cytosolic guanylate cyclase-bound heme to generate the NO-heme adduct of guanylate cyclase. NO can readily react with other cellular hemoproteins such as hemoglobin and myoglobin to produce corresponding NO-heme adducts that can rapidly activate

guanylate cyclase (9, 35, 36). Several in vitro studies directly established the role of NO in angiogenesis (72, 73). Treatment of HepG2 cells with NO donor SNAP was found to increase VEGF mRNA expression. Guanylate cyclase is likely to be important for NO-mediated VEGF activation (8). There is also considerable evidence that NO downregulates the expression of the VEGF gene (57, 63). Despite several negative observations, activation of angiogenesis in mammalian (human) monocytes is believed to be NO-dependent (30). Indeed, several studies have documented that NO-generating compounds stimulate angiogenesis in human glastoma and hepatoma cells (8). A positive correlation was found between nitric oxide synthase, cGMP levels, and tumor angiogenesis in head and neck cancer (18). However, the role of NO in angiogenesis is still controversial. For example, NO donors were found to inhibit angiogenic activity in the chick chorioallantoic membrane (45) and the growth and metastatic properties of Lewis lung tumor in mice (44). In addition, NO donors inhibit VEGF expression in the arterial wall in response to balloon angioplasty (63), and in rat lungs during acute and chronic hypoxia (5). Transfer of eNOS (66) and iNOS (65) genes resulted in the inhibition of restenosis after balloon angioplasty. In contrast, a recent study showed Larginine supplementation after balloon angioplasty of rabbit iliac arteries was beneficial for the healing of endothelium, demonstrating the positive role of NO in angiogenesis (52). Human colon cancer cell lines transfected with a NOS gene grow faster and become more vascularized than normal cell lines in vivo (24). NO also enhances the proliferation and migration of endothelial cells in vitro (72). Thus, a significant number of studies have demonstrated that NO may stimulate the proliferation of endothelial cells while others failed to prove that such a mechanism exists and suggested an inhibitory effect of NO on endothelial proliferation.

THIOREDOXIN-GLUTAREDOXIN REDOX SIGNALING IN VEGF EXPRESSION

Thioredoxin (Trx) appears to play a crucial role in the redox regulation of the ROS signaling during and/or following ischemia/reperfusion. This protein, Trx, is an important component of the cellular defense against cardiac injury. Oxidized thioredoxin was found to be released into the plasma of patients undergoing cardiopulmonary bypass surgery (42). It is one of the major cellular protein disulfide reductases ubiquitously present in mammalian tissues, including myocardium. They possess dithiol/disulfide active sites and can serve as electron donors for enzymes, including thioredoxin peroxidases and ribonucleotide reductases (2). Thioredoxins are critical for redox regulation of protein function and signaling via thiol-redox control. Thioredoxins are reduced by electrons from NADPH via thioredoxin reductase. Recent studies from our laboratory have provided evidence for redox regulation of myocardial ischemia/reperfusion (34). There are two major thioredoxins: Trx1, a cytosolic and nuclear form, and thioredoxin 2 (Trx2), a mitochondrial form. Trx1 is the major thioredoxin-redox protein, which is responsible for most of

the biological signals (22, 41). It plays a crucial role in thiolredox control of cell function through transcription regulation of target genes including that of NF-kB, which controls numerous gene expressions. Today the glutaredoxin and thioredoxin systems are considered to be parallel redox systems. In fact, the absence of cross-reactivity between the redoxins and the respective NADPH-dependent reductase may have special importance in regulation because the systems can operate independently. In mammalian cells, there seems to be cross talk between the thioredoxin system and the glutaredoxins (22). Very recently, we documented resveratrol-mediated induction of Trx-1, which shows sequential activation and expression of HO-1 as well as pro-angiogenic factor and cardioprotective molecule VEGF in both in vitro and in vivo models. We have shown that adjunctive treatment with SnPP significantly inhibits all the VEGF-induced angiogenic activities of resveratrol and Trx-1 in vitro and in vivo (Fig. 2) (26). Our study is in agreement with an earlier report showing that overexpression of HO-1 augments the angiogenic effect in endothelial cells (12) and the activation and overexpression of HO-1 leading to the upregulation of VEGF synthesis. We have explored resveratrol-mediated expression of Trx-1, showing cardioprotective effects in the myocardial infarction model with increased perfused capillary density in peri-infarct myocardium, along with improved cardiac function. The cardioprotective effect is significantly attenuated by SnPP (26). It was reported previously that the redox protein Trx-1 increases hypoxia inducible factor-1α (HIF-1α) protein expression under both normoxic and hypoxic conditions. This is found to be associated with augmented VEGF formation and increased tumor angiogenesis in vivo (67). The hypoxia inducible factor-1 complex influences the expression of many genes including VEGF (53). Thus, VEGF is implicated as a major angiogenic factor leading to the development of new vessels from pre-existing capillaries. Transfection of cells with human Trx-1 increases the overall production of VEGF in MCF-7

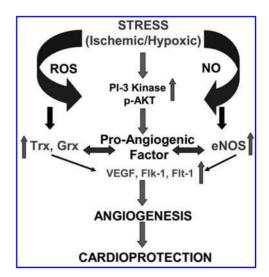


FIG. 2. Schematic diagram suggesting the involvement of reactive oxygen species and redox regulated protein Trx and Grx in angiogenesis, followed by myocardial protection.

breast cancer, HT-29 colon cancer, and WEHI7.2 lymphoma cells [68].

In summary, the angiogenic response in vascular tissue is triggered by ROS signaling in a highly coordinated manner. It appears that massive amounts of ROS produced during ischemia and reperfusion in the vascular tissue, especially in the heart, cause significant injury to cardiomyocytes and endothelial cells. However, during the reperfusion, the same ROS potentiate the repair process and trigger a signal transduction cascade leading to angiogenesis. Significant efforts in this area of research have led to the discovery of a growing number of pro- and anti-angiogenic molecules, some of which are already in clinical trials. However, there are several outstanding questions that must be addressed for successful translation of discoveries from the bench to the bedside. With advances in molecular genetics and the availability of molecular probes, imaging technologies, and therapeutic opportunities, we are now beginning to answer these questions.

ACKNOWLEDGMENTS

This study was supported by National Institutes of Health Grants HL 56803, HL 69910, and HL 85804.

ABBREVIATIONS

APC, adenomatous polyposis coli protein; H₂O₂, hydrogen peroxide; IPC, ischemic preconditioning; MI, myocardial infarction; ROS, reactive oxygen species; O₂-*, superoxide anions; VEGF, vascular endothelial cells.

REFERENCES

- Addya S, Shiroto K, Turoczi T, Zhan Lijun, Kaga S, Fukuda S, Surrey S, Duan Li–Juan, Fong G–H, Yamamoto F, and Maulik N. Ischemic preconditioning-mediated cardioprotection is disrupted in heterozygous Flt-1 (VEGFR-1) knockout mice. *J Mol Cell Car-diol* 38: 345–351, 2005.
- Amer ESJ and Holmgren A. Physiological functions of thioredoxin and thioredoxin reductase. Eur J Biochem 267: 6102–6109, 2000.
- Asahara T, Chen D, Takahashi T, Fujikawa K, Kearney M, Magner M, Yancopoulos GD, and Isner JM. Tie-2 receptor ligands, angiopoietin-1 and angiopoietin-2, modulate VEGF-induced postnatal neovascularization. *Circ Res* 83: 342–343, 1998.
- Babior BM. The NADPH oxidase of endothelial cells. *IUMB Life* 50: 267–269, 2000.
- Banai S, Jaklitsch MT, Shou M, Lazarous DF, Scheinowitz M, Biro S, Epstein SE, and Unger EF. Angiogenic-induced enhancement of collateral blood flow to ischemic myocardium by vascular endothelial growth factor in dogs. *Circulation* 89: 2183–2189, 1994.
- Banai S, Shweiki D, Pinson A, Chandra M, Lazarovici G, and Keshet E. Upregulation of vascular endothelial growth factor expression induced by myocardial ischemia: implications for coronary angiogenesis. *Cardiovasc Res* 28: 1176–1179, 1994.
- Brand T, Sharma HS, Fleischmann KE, Duncker DJ, McFalls EO, Verdue PD, and Schaper W. Proto-oncogene expression in porcine myocardium subjected to ischemia and reperfusion. *Circ Res* 71:1351–1360, 1992.
- Chin K, Kurashima Y, Ogura T, Tajiri H, Yoshida S, and Esumi H. Induction of vascular endothelial growth factor by nitric oxide in

- human glioblastoma and hepatocellular carcinoma cells. *Onco-gene* 15:437–442, 1997.
- Chinkers M and Garbers DL Signal transduction by guanylyl cyclases. Ann Rev Biochem 60: 553–575, 1991.
- Chua CC, Hamdy RC, and Chua BHL. Upregulation of vascular endothelial growth factor by H₂O₂ in rat heart endothelial cells. Free Rad Biol Med 25: 891–897, 1998.
- Crow JP. Dochlorodihydrofluorescein and dihydrorhodamine-123 are sensitive indicators of peroxynitrite *in vitro*: implications for intracellular measurement of reactive nitrogen and oxygen species. *Nitric Oxide* 1: 145–157, 1997.
- 12. Deramaudt BM, Braunstein S, Remy P, and Abraham NG. Gene transfer of human heme oxygenase into coronary endothelial cells potentially promotes angiogenesis. *J Cell Biochem* 68:121–127, 1008
- Donnini S and Ziche M. Constitutive and inducible nitric oxide synthase: role in angiogenesis. *Antioxid Redox Signal* 4: 817–823, 2002.
- Ellis EA, Guberski DL, Somogyi–Mann M, and Grant MB. Increased H2O2, vascular endothelial growth factor and receptors in the retina of the BBZ/Wor diabetic rat. Free Radic Biol Med 28: 91–101, 2000.
- Ely SW and Berne RM. Protective effects of adenosine in myocardial ischemia. *Circulation* 85: 893–904, 1992.
- Fong G-H, Rossant J, Gerstenstein M, and Breitman M L. Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium. *Nature* 376: 66-70, 1995.
- Fukuda S, Kaga S, Sasaki H, Zhan L, Zhu L, Otani H, and Maulik N. Angiogenesis signal triggered by ischemic stress induces myocardial repair in rat during chronic infarction. *J Mol Cell Cardiol* 36: 547–559, 2004.
- Gallo O, Masini E, Morbidelli L, Franchi A, Finistorchi I, Vergari WA, and Ziche M. Role of nitric oxide in angiogenesis and tumor progression in head and neck cancer. *J Natl Cancer Inst* 90: 587–596, 1998.
- Hakada H, Inagaki M, Kawamoto S, and Sasaki, Y. Isoquinoline sulfonamides, novel and potent inhibitors of cyclic nucleotide dependent protein kinase and protein kinase C. *Biochem* 23: 5036– 5041, 1984.
- Hartnett ME, Garcia CM, and D'Amore PA. Release of bFGF, an endothelial cell survival factor, by osmotic shock. *Invest Opthal*mol Vis Sci 40: 2945–2951, 1999.
- Hashimoto E, Ogita T, Nakaoka T, Matsuoka R, Takao A, and Kira Y. Rapid induction of vascular endothelial growth factor expression by transient ischemia in the rat heart. *Am J Physiol* 267: H1948–H1954, 1994.
- Holmgren A. Thioredoxin and glutaredoxin systems. J Biol Chem 264: 13963–13963, 1989.
- 23. Isner J M, Pieczek A, Schainfed R, Blair R, Haley L, Asahara T, Rosenfield K, Razvi S, Walsh K, and Symes J. Clinical evidence of angiogenesis is after arterial gene transfer of ph VEGF165 in patient with ischemic limb. *Lancet* 348: 370–374, 1996.
- Jenkins DC, Charles IG, Thomsen LL, Moss DW, Holmes LS, Baylis SA, Rhodes P, Westmore K, Emson PC, and Moncada S. Roles of nitric oxide in tumor growth. *Proc Natl Acad Sci USA* 92: 4392–4396, 1995.
- Jian Li, Brown LF. Hibberd MG. Grossman JD. Morgan JP, and Simon M. VEGF, Flk-1, Flt-1 expression in a rat myocardial infarction model of angiogenesis. *Am J Physiol* 270: H1803–H1811, 1996.
- Kaga S, Zhan I, Matsumoto M, and Maulik N. Resveratrol enhances neovascularization in the infracted rat myocardium through the induction of thioredoxin-1, heme oxygenase-1 and vascular endothelial growth factor. *J Mol Cell Cardiol* 39: 813–822, 2005.
- Kaipainen A, Korhonen J, Pajusola K, Aprelikova O, Persico MG, Terman BI, and Alitalo K. The related Flt-4, Flt-1, and KDR receptor tyrosine kinases show distinct expression patterns in human fetal endothelial cells. *J Exp Med* 178: 2077–2088, 1993.
- 28. Kwata H, Yoshida K-I, Kawamoto A, Kurioka H, Takase E, Sasaki Y, Hatanaka K, Kobayashi M, Ueyama T, Hashimoto T, and Dohi K. Ischemic preconditioning upregulates vascular endothelial growth factor mRNA expression and neovascularization via nu-

- clear translocation of protein kinase C epsilon in the rat ischemic myocardium. *Circ Res* 88: 696–704, 2001.
- 29. Ladoux A and Frelin C. Expression of vascular endothelial growth factor by cultured endothelial cells from brain microvessels. *Biochem Biophys Res Commun* 194: 799–803, 1993.
- Leibovich SJ, Polverini PJ, Fong TW, Harlow LA, and Koch A. Production of angiogenic activity by human monocytes requires an L-arginine/nitric oxide-synthase-dependent effector mechanism. *Proc Natl Acad Sci USA* 91: 4190–4194, 1994.
- 31. Lelkes PI, Hahn KL, Sukovich DA, Karmiol S, and Schmidt DH. On the possible role of reactive oxygen species in angiogenesis. *Adv Exp Med Biol* 454: 295–310, 1998.
- Li J, Brown LF, Hibberd MG, Grossman JD, Morgan JP, and Simons M. VEGF, Flk-1, and Flt-1 expression in a rat myocardial infarction model of angiogenesis. *Am J Physiol* 270: H1803–H1811, 1996
- 33. Maulik N and Das DK. Potentiation of angiogenic response by ischemic and hypoxic preconditioning of the heart. *J Cell Mol Med* 6: 13–24, 2002.
- 34. Maulik N and Das DK. Redox signaling in vascular angiogenesis. *Free Rad Biol Med* 33: 1047–1060, 2002.
- Maulik N, Engelman DT, Watanabe M, Engelman RM, Rousou JA, Flack JE, Deaton DW, Gorbunov NV, Elsayed NM, Kagan VE, and Das DK. Nitric oxide/carbon monoxide: a molecular switch for myocardial preservation during ischemia. *Circulation* 94: 398–406, 1996.
- Maulik N, Engelman DT, Watanabe M, Engelman RM, and Das DK. Nitric oxide–a retrograde messenger for carbon monoxide signaling in ischemic heart. *Mol Cell Biochem* 157: 75–86, 1996.
- Maulik N, Goswami S, Galang N, and Das DK. Differential regulation of Bcl-2, AP-1 and NF-κB on cardiomyocyte apoptosis during myocardial ischemic stress adaptation. FEBS Lett 443: 331–336, 1999.
- Maulik N. Redox regulation of vascular angiogenesis (forum editorial) Antioxid Redox Signal 4: 783–784, 2002.
- Maulik N. Redox signaling of angiogenesis. Antioxid Redox Signal 4: 805–815, 2002.
- Miller PD, Telford IR, and Haas GR. Effect of hyperbaric oxygen on cardiogenesis in the rat. Biol Neonate 17: 44–52, 1971.
- Nakamura H, Nakamura K, and Yodoi J. Redox regulation of cellular activation. *Anual Rev Immunol* 15: 351–369, 1997.
- Nakamura H, Vaage J, Valen G, Padilla CA., Bjornstedt M, and Holmgren A. Measurements of plasma glutaredoxin and thioredoxin in healthy volunteers and during open-heart surgery. Free Rad Biol Med 24: 1176–1186, 1998.
- Pearlman JD, Hibberd MG, Chuang ML, Harada K, Lopez JJ, Gladstone SR, Friedman M, Sellke FW, and Simons M. Magnetic resonance mapping demonstrates benefits of VEGF-induced myocardial angiogenesis. *Nat Med* 1: 1085–1089, 1995.
- 44. Pipili–Synetos E, Papa Georglou A, Sakkoula E, Sotiropoulou G, Fotsis T, Karakiulakis G, and Maragoudakis ME Inhibition of angiogenesis, tumour growth and metastasis by the NO-releasing vasodilators, isosorbide mononitrate and dinitrate. *Br J Pharmacol* 116: 1829–1834, 1995.
- Pipili–Synetos E, Sakkoula E, Haralabopoulos G, Andriopoulou P, Peristeris P, and Maragoudakis ME. Evidence that nitric oxide is an endogenous antiangiogenic mediator. *Br J Pharmacol* 111: 894–902, 1994.
- Quinn MT, Evans T, Loetterle LR, Jesaitis AJ, and Bokoch GM. Translocation of Rac correlates with NADPH oxidase activation. Evidence for equimolar translocation of oxidase components. *J Biol Chem* 268: 20983–2097, 1993.
- Ray PS, Estrada—Hernandez T, Sasaki H, Zhu Li, and Maulik N. Early effects of hypoxia/reoxygenation on VEGF, Ang-1, Ang-2 and their receptors in the rat myocardium: Implications for myocardial angiogenesis. *Mol Cell Biochem* 213: 145–153, 2000.
- Ray PS, Sasaki H, Estrada–Hernandez T, Zhu Li, and Maulik N. Effects of hypoxia/reoxygenation on angiogenic factors and their tyrosine kinase receptors in the rat myocardium. *Antoxid Redox* Signal 3: 89–102, 2001.

- Ruef J, Hu ZY, Yin LY, Wu Y, Hanson SR, and Kelly AB. Induction of vascular endothelial growth factor in balloon-injured baboon arteries. *Circ Res* 81: 24–33, 1997.
- Sasaki H, Fukuda S, Otani H, Zhu Li, Yamaura G, Engelman RM, Das DK, and Maulik N. Hypoxic preconditioning triggers myocardial angiogenesis: a novel approach to enhance contractile functional reserve in rat with myocardial infarction *J Mol Cell Cardiol* 34: 335–348, 2002.
- Sasaki H, Ray PS, Zhu Li, Otani H, Asahara T,and Maulik N. Hypoxia/reoxygenation promotes myocardial angiogenesis via an NFκB-dependent mechanism in a rat model of chronic myocardial infarction. *J Mol Cell Cardiol* 33: 283–294, 2001.
- Schwarzacher S, Lim TT, Wang BY, Kernoff RS, Niebauer J, Cooke JP, and Yeung AC. Local intramural delivery of L-arginine enhances nitric oxide generation and inhibits lesion formation after vascular injury. *Circulation* 95:1863–1869, 1997.
- Semenza GL. HIF-1: mediator of physiological and pathophysiological responses to hypoxia. J Appl Physiol 88: 1474–1480, 2000.
- Sen CK, Khanna S, and Roy S. Perceived hyperoxia: oxygen-induced remodeling of the reoxygenated heart. *Cardiovasc Res* 71: 280–288, 2006
- Shalaby F, Rossant J, Yamaguchi TP, Gertsenstein M, Wu XF, Breitman ML, and Schuh AC. Failure of blood-island formation and vasculogenesis in Flk-1 deficient mice. *Nature* 376: 62–66, 1995.
- 56. Shih S–C, Mullen A, and Abrams K, Mukhopadhyay D, and Claffey KP. Role of protein kinase C isoforms in phorbol ester-induced vascular endothelial growth factor expression in human glioblastoma cells. *J Biol chem* 274: 15407–15414, 1999.
- Sogawa K, Numayama Tsuruta K, Ema M, Abe M, Abe H, and Fuji Kuriyama Y. Inhibition of hypoxia-inducible factor 1 activity by nitric oxide donors in hypoxia. *Proc Natl Acad Sci USA* 95: 7368–7373, 1998.
- Spyridopoulos I, Brogi E, Kearney M, Sullivan AB, Cetrulo C, Isner JM, and Losardo DW. Vascular endothelial growth factor inhibits endothelial cell apoptosis induced by tumor necrosis factoralpha: balance between growth and death signals. *J Mol Cell Cardiol* 29: 1321–1330, 1997.
- Takeshita S, Zheng LP, Brogi E, Kearney M, and Pu LQ. Therapeutic angiogenesis. A single intra arterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. *J Clin Invest* 93: 662–670, 1994.
- 60. Takeya R, Ueno N, Kami K. Taura M, Kohjima M, Izaki T, Nunoi H, and Sumimoto H. Novel human homologues of p47phox and p67 phox participate in activation of superoxide-producing of NADPH oxidases. *J Biol Chem* 278: 25234–25246, 2003.
- Terman BI, Vermazen MD, Carrion ME, Dimitrov D, Armellino DC, Gospodarowicz D, and Bohlen P. Identification of KDR tyrosine kinase as a receptor for vascular endothelial cell growth factor. *Biochem Biophys Res Commun* 187: 1579–1586, 1992.
- Tojo T, Fukai MU, Tojo MY, Ikeda S, Patrushev N, and Alexander W. Role of gp91^{phox} (Nox2)-containing NAD(P)H oxidase in angiogenesis in response to hindlimb ischemia. *Circulation* 111: 2347–2355, 2005.
- Tsurumi Y, Murohara T, Krasinski K, Chen D, Witzenbichler B, Kearney M, Couffinhal T, and Isner JM. Reciprocal relation between VEGF and NO in the regulation of endothelial integrity. *Nat Med* 3: 879–886, 1997.
- 64. Tuder RM. Voelkel NF, and Flook BE. Increased gene expression for VEGF and the VEGF receptors KDR/Flk and Flt in lungs exposed to acute or to chronic hypoxia. Modulation of gene expression by nitric oxide. *J Clin Invest* 95:1798–1807, 1995.
- Tzeng E, Shears LL, Robbins PD, Pitt BR, Geller DA, Watkins SC, Simmons Rl, and Billiar TR. Vascular gene transfer of the human inducible nitric oxide synthase characterization of activity and effects of myointimal hyperplasia. *Mol Med* 2: 211–225, 1996.
- 66. Von der Leyen HE, Gibbons GH, Morishita R, Lewis NP, Zhang L, Nakajima M, Kaneda Y, Cooke JP, and Dzau VJ. Gene therapy inhibiting neointimal vascular lesion: in vivo transfer of endothelial cell nitric oxide synthase gene. Proc Natl Acad Sci USA 92: 1137–1141, 1995.
- Welsh SJ, Bellamy WT, Briehl MM, and Powis G. The redox protein thioredoxin-1 (Trx-1) increases hypoxia-inducible factor

1alpha protein expression: Trx-1 overexpression results in increased vascular endothelial growth factor production and enhanced tumor angiogenesis. *Cancer Res* 62: 5089–5095, 2002.

- 68. Welsh SJ, Williams RR, Birmingham A, Newman DJ, Kirkpatrick DL, and Powis G. The thioredoxin redox inhibitors 1-methyl-propyl 2- imidazolyl disulfide and pleurotin inhibit hypoxia induced factor 1 and vascular endothelial growth factor formation. Mol Cancer Ther 2: 235–243, 2003.
- Witzenbichler B, Maisonpierre P C, Jones P, Yancopoulos GD, and Isner JM. Chemotactic properties of angiopoietin-1, angiopoietin-2, ligands for the endothelial-specific receptor tyrosine kinase Tie2. *J Biol Chem* 273: 18514–18521, 1998.
- Woldback PR, Hoen, IB, Christensen G, and Tonnessen T. Gene expression of colony-stimulating factors and stem cell factor after myocardial infarction in the mouse. *Acta Physiol Scand* 175: 173–181, 2002.
- Zhu Li, Fukuda S, Das DK, and Maulik N. Anti-apoptotic protein survivine plays a significant role in tubular morphogenesis of human arteriolar endothelial cells by hypoxic preconditioning. FEBS Lett 508: 369–374, 2001.
- Ziche M, Morbidelli L, Masini E, Amerini S, Granger HJ, Maggi CA, Geppetti P, and Ledda F. Nitric oxide mediates angiogenesis

- *in vivo* and endothelial cell growth and migration *in vitro* promoted by substance P. *J Clin Invest* 94: 2036–2044, 1994.
- Ziche M, Morbidelli L, Choudhuri R, Zhang HT, Donnini S, Granger HJ, and Bicknell R. Nitric oxide synthase lies downstream from vascular endothelial growth factor-induced but not fibroblast growth factor induced angiogenesis. *J Clin Invest* 99: 2625–2634, 1997.

Address reprint requests to:
Nilanjana Maulik
Molecular Cardiology and Angiogenesis Laboratory
Department of Surgery
University of Connecticut School of Medicine
Farmington, CT 06030–1110

E-mail: nmaulik@neuron.uchc.edu

Date of first submission to ARS Central, July 7, 2006; date of acceptance, July 8, 2006.

This article has been cited by:

- Mohamed B. Khadeer Ahamed, Abdalrahim F. A. Aisha, Zeyad D. Nassar, Jamshed M. Siddiqui, Z. Ismail, Sms Omari, Cr Parish, A. M. S. Abdul Majid. 2011. Cat's Whiskers Tea (Orthosiphon Stamineus) Extract Inhibits Growth of Colon Tumor in Nude Mice and Angiogenesis in Endothelial Cells via Suppressing VEGFR Phosphorylation. *Nutrition and Cancer* 111202073727007. [CrossRef]
- 2. Antonio D. Lassaletta, Louis M. Chu, Frank W. Sellke. 2011. Therapeutic neovascularization for coronary disease: current state and future prospects. *Basic Research in Cardiology*. [CrossRef]
- 3. Gel R.M. Berardi, Carmen K. Rebelatto, Heloísa F. Tavares, Max Ingberman, Patrícia Shigunov, Fabiane Barchiki, Alessandra M. Aguiar, Nelson I. Miyague, Julio C. Francisco, Alejandro Correa. 2011. Transplantation of SNAP-treated adipose tissue-derived stem cells improves cardiac function and induces neovascularization after myocardium infarct in rats. *Experimental and Molecular Pathology* **90**:2, 149-156. [CrossRef]
- 4. Ramón Rodrigo, Andrés Miranda, Leonardo Vergara. 2011. Modulation of endogenous antioxidant system by wine polyphenols in human disease. *Clinica Chimica Acta* **412**:5-6, 410-424. [CrossRef]
- 5. Olivier Blanc-BrudeMyocardial Cell Death and Regeneration 66-80. [CrossRef]
- 6. Ligang Zhou, Rita Yn Tz Sung, Karen Li, Nga Hin Pong, Ping Xiang, Jie Shen, Pak Cheung Ng, Yuan Chen. 2010. Cardioprotective effect of dexrazoxane in a rat model of myocardial infarction: Anti-apoptosis and promoting angiogenesis. *International Journal of Cardiology*. [CrossRef]
- 7. Masuko Ushio-Fukai Approaches for Studying Redox Signaling in Angiogenesis 191-200. [Abstract] [Summary] [Full Text PDF] [Full Text PDF with Links]
- 8. Lindsay Brown, Paul A. Kroon, Dipak K. Das, Samarjit Das, Arpad Tosaki, Vincent Chan, Manfred V. Singer, Peter Feick. 2009. The Biological Responses to Resveratrol and Other Polyphenols From Alcoholic Beverages. *Alcoholism: Clinical and Experimental Research* 33:9, 1513-1523. [CrossRef]
- 9. Munir Boodhwani, Frank W. Sellke. 2009. Therapeutic Angiogenesis in Diabetes and Hypercholesterolemia: Influence of Oxidative Stress. *Antioxidants & Redox Signaling* 11:8, 1945-1959. [Abstract] [Full Text PDF] [Full Text PDF] with Links]
- 10. L. F. Wetherill. 2009. Can ESR be used to assess the levels of oxidative stress in fat-loaded human hepatocytes and hepatic stellate cells?. *Bioscience Horizons* **2**:2, 197-204. [CrossRef]
- 11. C SEN, S ROY. 2008. Redox signals in wound healing. *Biochimica et Biophysica Acta (BBA) General Subjects* **1780**:11, 1348-1361. [CrossRef]
- 12. Suwan Yap, Karen J. Loft, Owen L. Woodman, Spencer J. Williams. 2008. Discovery of Water-Soluble Antioxidant Flavonols without Vasorelaxant Activity. *ChemMedChem* 3:10, 1572-1579. [CrossRef]
- 13. Zita Hertelendi, Attila Tóth, Attila Borbély, Zoltán Galajda, Jolanda van der Velden, Ger J.M. Stienen, István Édes, Zoltán Papp. 2008. Oxidation of Myofilament Protein Sulfhydryl Groups Reduces the Contractile Force and Its Ca2+ Sensitivity in Human Cardiomyocytes. *Antioxidants & Redox Signaling* 10:7, 1175-1184. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 14. Masuko Ushio-Fukai, Yoshimasa Nakamura. 2008. Reactive oxygen species and angiogenesis: NADPH oxidase as target for cancer therapy. *Cancer Letters* **266**:1, 37-52. [CrossRef]
- 15. Yasar Kucukardali, Sebnem Aydogdu, Namik Ozmen, Arif Yonem, Emrullah Solmazgul, Mustafa Ozyurt, Yilmaz Cingozbay, Aydogan Aydogdu. 2008. The relationship between severity of coronary artery disease and plasma level of vascular endothelial growth factor. *Cardiovascular Revascularization Medicine* 9:2, 66-70. [CrossRef]
- 16. Sashwati Roy, Savita Khanna, Chandan K. Sen. 2008. Redox regulation of the VEGF signaling path and tissue vascularization: Hydrogen peroxide, the common link between physical exercise and cutaneous wound healing. *Free Radical Biology and Medicine* **44**:2, 180-192. [CrossRef]
- 17. Gitika KHANNA, Vishal DIWAN, Manjeet SINGH, Nirmal SINGH, Amteshwar S. JAGGI. 2008. Reduction of Ischemic, Pharmacological and Remote Preconditioning Effects by an Antioxidant N-Acetyl Cysteine Pretreatment in Isolated Rat Heart. *YAKUGAKU ZASSHI* 128:3, 469-477. [CrossRef]
- 18. J Zargooshi. 2007. Open prostatectomy for benign prostate hyperplasia: short-term outcome in 3000 consecutive patients. *Prostate Cancer and Prostatic Diseases* **10**:4, 374-377. [CrossRef]
- 19. Beatrix Feigl. 2007. Age-related maculopathy in the light of ischaemia. *Clinical and Experimental Optometry* **90**:4, 263-271. [CrossRef]

- 20. Masuko Ushio–Fukai . 2007. VEGF Signaling Through NADPH Oxidase-Derived ROS. *Antioxidants & Redox Signaling* 9:6, 731-739. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 21. Dipak K. Das Methods in Redox Signaling . [Citation] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]